

## Lymphocytes purification from cattle and caprine blood samples - May 28th 2015

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### REAGENTS/ SOLUTIONS:

PBS/EDTA (= PBS without Ca and Mg (lab made) with EDTA 2 mM)(ref Sigma E5134)

Histopaque-1077 (Sigma-Aldrich ref 10771)

PBS/EDTA/FCS(= PBS with EDTA 2 mM +5% FCS + (optional) 1% P-S (= Penicillin-Streptomycin)

Lysis buffer (= NH<sub>4</sub>Cl 155mM + KHCO<sub>3</sub> 10mM + EDTA 0.2 mM)

RPMI 1640 with L-glutamine and NaHCO<sub>3</sub> ref Sigma-Aldrich R8758

RPMI/FCS (with 10% FCS)

RPMI/DMSO (= RPMI with 10% DMSO ref Sigma D2650)

### EQUIPMENT:

50ml polypropylene tubes

Centrifuge

1.8 ml cryotubes

Cell freezing container CoolCell®LX (BioCision)

### PROCESS

#### 1. Dilution of the blood:

Prepare 50ml polypropylene tubes containing 25 ml PBS EDTA/ 2mM buffer. Dilute the collected blood **at least to half** by pouring 25 ml blood into the buffer. Homogenize gently.

#### 2. Purification of Mononuclear cells:

- a. Distribute gently 30-35 mL of diluted blood over 15ml Histopaque-1077 layer without mixing the two layers
- b. Spin at 1500g for 30', at room temperature (set the lowest acceleration and deceleration of the centrifuge).
- c. Transfer the PBMCs layer into a 50ml polypropylene tube filled with 10 ml of PBS/EDTA/FCS. Fill tube up to 50ml with PBS/EDTA/FCS medium.
- d. Spin at 450/500g for 10', at 4°C
- e. Resuspend cells in PBS/EDTA/FCS
- f. Spin at 400g for 10', at 4°C

#### 3. Red blood cell lysis:

- a. Add 7 ml of lysis buffer on each pellet. After 2 minutes, stop the lysis by adding RPMI10%FCS medium up to 45 ml (this step is to remove red blood cells)
- b. Spin at 400g for 10', at 4°C

- c. Resuspend the cell pellet in 10 ml RPMI/10%FCS medium and count cells
- b. Spin at 400 g for 10', at 4°C.

**4. Cell freezing:**

- a. Resuspend  $10^8$  PBMCs in 1.8 ml of cold FCS /DMSO in cryotubes (work on ice).
- b. Transfer cryotubes to a cell freezing container and place at -80°C
- c. About 48 to 72 hours later, transfer cryotubes to a liquid nitrogen container for long term storage.